

# LABQ TAQ DNA POLYMERASE

LabQ Taq DNA Polymerase is a robust and reliable Taq DNA polymerase suitable for all common PCR applications like colony PCR, cloning applications, high-throughput PCR and routine PCR.

## **STORAGE CONDITIONS**

Store all components at -20°C and avoid repeated freeze and thaw cycles.

#### **MATERIALS PROVIDED**

100  $\mu$ l LabQ Taq DNA Polymerase 5 U/ $\mu$ l (green cap) 2x 1.5 ml 10X PCR Buffer (blue cap)

### **ADDITIONAL MATERIALS REQUIRED**

Nuclease free dH<sub>2</sub>O Nuclease free PCR tubes / plates & sealing options Thermocycler PCR Primer (10  $\mu$ M each) dNTP Mix (10 mM each) template DNA

#### **REACTION SETUP**

1) Thaw all reaction components completely and mix gently to ensure even distribution off all components. Prepare the reaction on ice in a sterile, nuclease free tube and mix gently after addition of the polymerase. Collect all liquid at the bottom of the tube by a quick spin.

COMPONENT	Volume	FINAL CONCENTRATION
10X PCR Buffer	5 μΙ	1X
dNTP Mix (10 mM each)	0.5 μΙ	0.1 mM each
Primer 1 (10 μM)	1 μΙ	$0.1 \ \mu M - 0.5 \ \mu M$
Primer 2 (10 μM)	1 μΙ	$0.1 \ \mu M - 0.5 \ \mu M$
LabQ Taq DNA Polymerase (5 U/μL)	0.2 μΙ	1 U
template DNA	1 μΙ	<1 μg
dH <sub>2</sub> O	to 50 μl	

2) Keep the reactions on ice until transfer to the thermocycler, then cycle according to these guidelines:

STEP	CYCLES	TEMPERATURE	DURATION
Initial Denaturation	1	94°C	5 minutes
		94°C	30 seconds
Amplification	30-35	$T_m - 5^{\circ}C$	30 seconds
		72°C	1 minute / kb
Final Extension	1	72°C	5 minutes
Hold	1	4°C	

3) Analyse the amplification reaction by gel electrophoresis using an acrylamide or agarose gel of appropriate percentage.